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Synthetic Studies on the Azinothricin Family of Antitumour Antibiotics. 5. Asymmetric Synthesis of Two Activated Esters For the Northern Sector of A83586C

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Abstract: An asymmetric synthesis of the activated esters 20 and 21 is reported. Both molecules equate with the C(28)-C(47) region of A83586C, and contain functionality appropriate for future unification with the hexapeptide unit. A useful new reaction is described for converting a methyl ester directly into a thioethyl or HOBT ester that operates under very mild conditions. Copyright © 1996 Elsevier Science Ltd

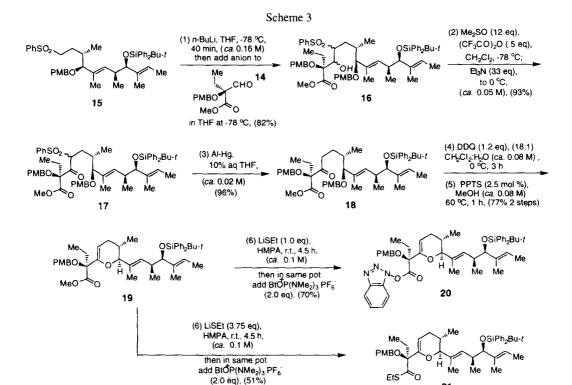
A83586C is an architecturally novel cyclodepsipeptide isolated from the antibiotic beer of *Streptomyces karnatakensis* by Smitka and coworkers. Although A83586C is itself highly toxic to mice at doses of 9.3 mg/kg, and unlikely ever to be used clinically, it remains an interesting drug design lead for the following reasons. First, it has an unprecedented molecular structure; second, it exhibits pronounced antitumour properties *in vitro* against a CCRF-CEM human T-cell leukaemia line (IC₅₀ = 0.0135 μg/ml), and; third, it has a devastating biocidal profile against resistant strains of Gram-positive bacteria at very low drug concentrations. Moreover, A83586C has a sister molecule, citropeptin, which can confer 123% life-extension on mice bearing P388 lymphocytic leukaemia when administered at 2 mg/kg/day,² although high toxicity is again a problematic feature. If the structural domains responsible for the observed toxicity of A83586C and citropeptin could be identified, through detailed structure-toxicity work, then it might prove possible to delete or modify these regions so as to create new analogues with enhanced antitumour effects and much lower toxicity *in vivo*. One way of gaining access to such analogues would be to develop a total synthesis of A83586C itself and then to make adjustments as required.³

Scheme 1

Some time ago, we reported the preparation of β -keto ester 3, an advanced intermediate with a structure corresponding to the C(28)-C(47) segment of the target molecule.^{3d} While it transpired that β -keto ester 3 could be readily manipulated into glycoside 2 (Scheme 1), subsequent elaboration of 2 into acid 1 proved troublesome.⁴ More specifically, we were unable to devise conditions for the successful hydrolysis of the *t*-butyl 1,3-dioxolane ester in 2 to obtain 1. In light of this set-back, we reluctantly modified our strategy to the northern half of A83586C, and now describe a route that provides the activated esters 20 and 21, two northern sector intermediates with functionality appropriate for unification with the hexapeptide sequence.

The starting compound for our synthesis of **20** and **21** was alkene **6**. It was subjected to Sharpless asymmetric dihydroxylation⁵ with AD-mix-β under the standard conditions to give diol **7** in 70-99% yield and 91% ee.⁶ Treatment of **7** with PPTS (2.5 mol%) and *p*-anisaldehyde dimethylacetal (1.2 eq) in dry DMF at 55 °C at 25 mm Hg for 2 h furnished **8** as a mixture of diastereoisomers in 98% yield. The latter were then reduced with *i*-Bu₂AlH⁷ at low temperature to give a 1:2 mixture of **9** and **10** in which the latter regioisomer predominated. Both alcohols were readily separated by flash chromatography. While it proved relatively straightforward to oxidise the primary alcohol in **10** to the corresponding carboxylic acid **11** with pyridinium dichromate in DMF, ⁸ the reaction did prove rather slow, taking **4** days to reach completion. Acid **11** was then

transformed into ester 12 by treatment with ethereal diazomethane in CHCl₃; the overall yield for the two steps was 64%. Esterification was followed by O-desilylation to obtain alcohol 13 and subsequent oxidation⁹ to aldehyde 14. The latter condensed readily with the α -phenylsulfonyl anion derived from 15^{3b} to give a mixture of β -hydroxy sulfones 16 that underwent Swern oxidation^{10,3b} to afford the β -keto sulfones 17 in 76% combined yield (Scheme 3). Desulfonylation of 17 proceeded efficiently with aluminium amalgam in aqueous THF¹¹ to produce ketone 18 in 96% yield. Treatment of 18 with 1.2 equiv. of DDQ¹² in aq. CH₂Cl₂ at 0 °C for 3 h brought about a remarkably regioselective O-debenzylation at C(34) to furnish an α/β mixture of ring-closed hemiketals in addition to hydroxy ketone. Exposure of this mixture to catalytic PPTS and methanol at 60 °C furnished glycal 19 in excellent yield. Unfortunately, all our attempts to directly hydrolyse the methyl ester in 19 with base to obtain the carboxylic acid were unsuccessful. Eventually, it was discovered that compound 19 could be converted directly into the activated N-hydroxybenzotriazolyl ester 20 by sequential dealkylative cleavage with LiSEt (1.0 eq) in HMPA¹³ for 4.5 h and addition of excess BOP reagent [benzotriazol-1-yloxy-tris(dimethyl-



amino)phosphonium hexafluorophosphate]¹⁴ to the reaction mixture. It also proved possible to convert 19 directly into the activated thioester 21 by reaction of the methyl ester with excess LiSEt in HMPA¹³ followed by addition of the BOP reagent. In our view, this new one-pot method for converting methyl esters directly into activated esters merits further attention since it operates under very mild conditions and should be applicable to other molecules in which conventional ester hydrolysis presents difficulties.

In summary, we have completed an asymmetric synthesis of the activated esters 20 and 21;15 both molecules have structures that are functionally equivalent to the C(28)-C(47) sector of A83586C. Further elaboration of 20 and 21 into A83586C and its analogues should now prove possible.

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- 15. All new compounds gave satisfactory IR, 400 MHz ¹H and 100 MHz ¹³C NMR spectra as well as HRMS and/or microanalyses within 0.4%. Selected physical data now follow (N.B. All ¹H and ¹³C NMR δ values are quoted in CDCl₃ as solvent and are relative to the residual CHCl₃ peak at 7.24 or 77.0 ppm respectively). Compound 18: 400 MHz ¹H NMR & 7.68-7.56 (m, 4H), 7.43-7.24 (m, 8H), 7.10 (m, 2H), 6.84 (m, 2H), $6.77 \text{ (m, 2H)}, 4.96 \text{ (m, 1H)}, 4.84 \text{ (d, } J = 9.9 \text{ Hz, 1H)}, 4.37 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H)}, 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1H}), 4.28 \text{ (1/2 AB q, } J = 10.7 \text{ Hz, 1$ J = 10.7 Hz, 1H), 4.14 (1/2 AB q, J = 11.3 Hz, 1H), 3.84 (1/2 AB q, J = 11.3 Hz, 1H), 3.78 (d, J = 8.7Hz, 1H), 3.77 (s, 3H), 3.74 (s, 3H), 3.66 (s, 3H), 3.00 (d, J = 9.6 Hz, 1H), 2.74-2.53 (m, 3H), 2.17 (m, 1H), 2.06 (m, 1H), 1.95 (m, 1H), 1.57 (m, 1H), 1.50 (m, 3H), 1.46 (s, 3H), 1.23 (d, J = 6.7 Hz, 3H), 1.21 (m, 1H), 1.04 (s, 9H), 0.91 (d, J = 6.7 Hz, 3H), 0.80 (t, J = 7.5 Hz, 3H), 0.54 (d, J = 6.7 Hz, 3H); 100 MHz ¹³C NMR δ 208.2, 169.9, 159.2, 158.9, 136.8, 136.3, 136.1, 134.5, 134.2, 133.6, 131.7, 131.0, 129.8, 129.5, 129.3, 129.9, 127.3, 127.0, 122.5, 113.7, 113.6, 89.8, 84.5, 69.0, 66.5, 55.3, 55.21, 55.19, 52.4, 37.6, 36.5, 34.4, 27.2, 26.9, 24.5, 19.6, 18.3, 15.9, 12.8, 11.1, 10.5, 7.3. Compound 19: $[\alpha]_D$ +29.5 ° (c 3.0 CH₂Cl₂); 100 MHz ¹³C NMR δ 172.3, 158.9, 150.7, 136.2, 136.1, 134.6, 134.3, 132.4, 131.6, 131.0, 129.3, 129.0, 127.2, 127.1, 122.1, 113.5, 97.9, 87.6, 83.9, 83.8, 66.1, 55.3, 51.9, 37.5, 29.1, 28.7, 27.2, 25.7, 19.6, 17.7, 17.4, 12.6, 11.7, 11.2, 7.7. Compound 20: $[\alpha]_D$ +35.0 ° (c 1.0 CH₂Cl₂); 400 MHz ¹H NMR δ 8.04 (d, J = 8.3 Hz, 1H), 7.65-7.56 (m, 4H), 7.43-7.24 (m, 11H), 6.84 (m, 2H), 5.23 (m, 1H), 5.04 (d, J = 9.0 Hz, 1H), 4.97 (m, 1H), 4.64(1/2 AB q, J = 10.5 Hz, 1H), 4.50 (1/2 AB q, J = 10.5 Hz, 1H), 3.82 (d, J = 7.6 Hz, 1H), 3.77 (s, 3H),3.63 (d, J = 9.5 Hz, 1H), 2.64 (m, 1H), 2.20 (m, 3H), 1.84 (m, 2H), 1.61 (s, 3H), 1.41 (s, 3H), 1.23 (d, J = 6.8 Hz, 3H), 1.03 (s, 9H superimposed on t, 3H), 0.92 (d, J = 6.7 Hz, 3H), 0.76 (d, J = 6.4 Hz, 3H); HRMS Calcd. for C₅₀H₆₁N₃O₆SiNa m/e 850.4227 (M+Na)+. Found: 850.4222. Compound 21: 400 MHz ¹H NMR δ 7.68-7.57 (m, 4H), 7.42-7.25 (m, 8H), 6.85 (d, J = 8.7 Hz, 2H), 4.96 (m, 2 H), 4.89 (d, J = 10.0 Hz, 1H), 4.43 (1/2 AB q, J = 10.7 Hz, 1H), 4.29 (1/2 AB q, J = 10.7 Hz, 1H)

1H), 3.78 (s, 3H superimposed on d, 1H), 3.51 (d, J = 9.0 Hz, 1H), 2.76 (m, 2H), 2.61 (m, 1H), 2.05 (m, 1H), 1.96 (m, 2H), 1.70 (m, 2H), 1.48 (s, 3H), 1.40 (s, 3H), 1.28 (d, J = 6.6 Hz, 3H), 1.18 (t, J = 7.4 Hz, 3H), 1.04 (s, 9H), 0.87 (d, J = 6.7 Hz, 3H), 0.80 (t, J = 7.3 Hz, 3H), 0.67 (d, J = 6.1 Hz, 3H);

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HRMS Calcd. for C₄₆H₆₂O₅SSiNa *m/e* 777.3985 (M+Na)+. Found: 777.3980.